

Dental microwear profilometry of African non-cercopithecoid catarrhines of the Early Miocene

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INTRODUCTION

The early Miocene of eastern Africa was home to a number of primate taxa. Their phylogenetic relationships and ecological niches have been much discussed, but not often agreed upon. Among the taxa present in these areas are two commonly recognized superfamilies, Proconsuloidea and Dendropithecoidea, along with a number of genera of uncertain taxonomic affiliation (*e.g.*, *Limnopithecus*, *Kalepithecus*.) ^{1, 2}. These primates existed from approximately 23 Ma - 16 Ma and represent a sizable part of the east African Miocene fossil record.

Previous analyses of these primates have focused on cladistics^{3,4}, postcranial functional morphology ^{5,6,7}, and dental morphology ^{8,9,10,11,12}. A less thoroughly explored area is the direct analysis of diet as a proxy for paleohabitat and paleobiology.

Here we examine the diets of east African Miocene non-cercopithecoid primates using dental microwear texture analysis, which has been widely used to reconstruct the diets and paleohabitats of a broad range of taxa ^{13,14,15,16,17}.

For this study we have assembled the largest microwear database of African Miocene catarrhines to date (n = 83), and have included *Limnopithecus*, a genus not analyzed in previous studies. This is the first direct comparison of the microwear <u>within the African Miocene catarrhines</u>, although some fossils have been used as comparative material for broader studies of Miocene primate paleoecology and paleobiology ¹⁸.

MATERIALS AND METHODS

Post-canine teeth for all available Early Miocene non-cercopithecoid catarrhine primates with potential microwear were collected from the Kenya National Museum in Nairobi, Kenya following standardized protocols ^{15,19,20}.

Genus	Dendropithecus	Limnopithecus	Micropithecus	Proconsul	Rangwapithecus		
Dendropithecus		0.942568	0.359036	0.810114	0.61569		
Limnopithecus			0.15656	0.49194	0.469933		
Micropithecus				0.257787	0.697534		
Proconsul					0.616872		
Rangwapithecus							
Wilks' lambda	a: 0.7	277	Pillai trace:		0.2985		
df1:	ź	24			24		
df2:	24	248.9			296		
F:	0.9	0.9894			0.0047		
p(same):	0.4	0.4808			0.3347		
Figure 4	. 01	0 1968			0.4731		
Eigenvalue 1		0.1500			57.98		
Eigenvalue 2	: 0.03	8411	Percent:		24.78		

 Table 1 – MANOVA table describing comparisons between fossil genera. P=0.05

Alouatta	3.48E ⁻⁰⁵ *	4.21E ^{-08*}	4.10E ⁻⁰³ *	1.26E ⁻⁰⁶ *	1.05E ⁻⁰⁴ *
Ateles	0.111	0.003*	0.175	0.226	0.042*
Cebus	2.91E ^{-04*}	2.16E ⁻⁰⁸ *	0.009*	1.63E ⁻⁰⁴ *	1.66E ⁻⁰⁵ *
Cercocebus	0.001*	1.23E ⁻⁰⁷ *	0.009*	0.002*	3.26E ⁻⁰⁵ *
Colobus	0.036*	2.13E ^{-04*}	0.004*	0.009*	0.001*
Macaca	0.311	0.014*	0.117	0.206	0.017*
Papio	0.020*	5.18E ⁻⁰⁵ *	0.042*	0.048*	0.001*
Presbytis	0.126	0.011*	0.390	0.069	0.159
Procolobus	0.025*	2.00E ⁻⁰⁴ *	0.021*	0.017*	0.002*
emnopithecus	0.019*	0.001*	0.199	0.003*	0.011*
heropithecus	0.013*	1.28E ⁻⁰⁴ *	0.033*	0.001*	0.002*
rachypithecus	0.146	0.028*	0.218	0.019*	0.055

Dendropithecus Limnopithecus Micropithecus Proconsul Rangwapithecus

0.126

0.022*

0.200

2.88E^{-04*}

0.129

0.254

0.511

0.091

0.744

0.165

0.523

0.021*

0.077

0.037*

0.269

0.004*

Table 2 – MANOVA table describing comparisons between fossil and extant genera. P=0.05. Significance indicated by *

DISCUSSION

• Results suggests that these fossil taxa were filling the common primate niche of generalized (and perhaps opportunistic) frugivory, though some differentiation may have existed (Figure 7).

Genus

Gorilla

Pan

Pongo

Lophocebus

0.779

0.165

0.705

0.062

Phase II crushing/grinding facets were inspected with a Sensofar PL μ confocal imaging profiler (Solarius, Inc) 100x objective lens. Point clouds were generated from 4 adjacent fields of 102 x 138 μ m with a lateral sampling interval of 0.18 μ m and vertical resolution of 0.005 μ m.

Point clouds were analyzed using scale-sensitive fractal analysis ²¹. This technique uses fractal geometry to analyze 6 variables to describe surface texture: Area scale fractal complexity (*Asfc*), Scale of maximum complexity (*Smc*), Anisotropy (*epLsar*), Textural fill volume (*Tfv*), and Heterogeneity (*HAsfc*) based on two grid densities, 3 x 3 (*HAsfc*₉) and 9x9 (*HAsfc*₈₁). See Figures 1 and 2 for graphical depictions of texture surfaces.

Microwear texture variables were collected from 83 individuals, encompassing 7 genera. Texture attributes were rank transformed and compared using conservative non-parametric statistical tests. Pairwise multivariate analysis of variance (MANOVA) tests were run within the fossil sample. An additional MANOVA was performed to compare 5 fossil genera (taxa with sample sizes >1)with a diverse group of extant primates from Scott *et al.*, 2012 ²⁰. Principal components analysis (PCA) was used for visualization co-variation and correlation of variables in n-dimensional space.



Figure 1 – Diagrammatic rendering of microwear surfaces

Figure 2 – Photosimulations of microwear surfaces

• The possibility of exogenous grit (e.g., dirt, ash, or other wind blown abrasives) affecting the microwear signature of the primate fossils is unlikely, as a recent study of fossil Tragulidae (*Dorcatherium*), from the same deposits indicate clear distinctions among 4 species ²⁴ (Figure 4).

• While no significant differences were seen among the fossils, PCA analyses show a possible dietary restriction of *Proconsul* on Songhor (possibly due to competition with *Rangwapithecus?*) (Figures 5 and 6).



Figure 3 – PCA of all fossil taxa. Significant overlap suggests no statistical difference in microwear texture variables.

Figure 4 – PCA plot showing separation of species from Rusinga and Songhor Island <u>Tragulidae (*Dorcatherium*)</u>²⁴.

Figure 5 – PCA of *Proconsul* microwear texture variables on Rusinga and Songhor.

Figure 6 – PCA of *Proconsul* and *Rangwapithecus* microwear texture variables on Rusinga and Songhor.

modified from Scott *et al.*, 2006²². Fields depict: 1. anisotropic texture, 2. complex texture, 3. heterogeneous texture, 4. homogeneous texture.

depicting examples of different texture types. Fields depict: 1. anisotropic texture, 2. complex texture, 3. heterogeneous texture, 4. homogeneous texture.

RESULTS

• Statistical tests show no significant difference among any of the fossil primates (Table 1), although there are significant differences between fossil microwear variables for the fossil taxa and those of extant primates (from Scott *et al.*, 2012)²⁰ (Table 2).

– Fossil genera are more commonly different from extant specialist feeders than from generalists

 Fossil genera are more commonly different from Platyrrhini and Cercopithecoidea than they are from Hominoidea.

•PCA/CVA analyses fail to show consistent differences in the morphospace of microwear texture variables in fossil genera and species

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REFERENCES

1. Harrison T. 2002. Late Oligocene to middle Miocene catarrhines from Afro-Arabia. In: Hartwig, WC (ed.), The Primate Fossil Record. Cambridge University Press, Cambridge. 2. Harrison T. 2010. Dendropithecoidea Proconsuloidea and Hominoidea. In: Werdelin, L. & Sanders, WJ (ed.), Cenozoic Mammals of Africa, Berkeley: University of California Press. 3. Andrews P. 1978. A revision of the Miocene Hominoidea of East Africa. Bulletin of the British Museum, Natural History, Geology, 30: 85–224. 4. Harrison T. 1987. The phylogenetic relationships of the early catarrhine primates: A review of the current evidence. Journal of Human Evolution, 16: 41–80. 5. Fleagle JG. 1983. Locomotor adaptation of Oligocene and Miocene hominoids and their phyletic implications. In: Ciochon RL and Corruccini R (eds.), New Interpretations of Ape and Human Ancestry. Plenum Press, New York. 6. Rose MD. 1983. Miocene Hominoid Postcranial Morphology: Monkey-like, Ape-like, Neither, or Both? In: Ciochon RL and Corruccini R (eds.), New Interpretations of Ape and Human Ancestry. Plenum Press, New York. 7. McHenry HM and Corruccini RS. 1983. The Wrist of Proconsul africanus and the Origin of Hominoid Postcranial Adaptations. In: Ciochon RL and Corruccini R (eds.), New Interpretations of Ape and Human Ancestry. Plenum Press, New York. 8 Gregory WK. 1922. Origin and Evolution of the Human Dentition. Williams & Wilkins Company, Baltimore, MD. 9. Kay RF, and Hiiemae KM. 1974. Jaw movement and tooth use in recent and fossil primates. Am. J. Phys. Anthropol. 40: 227-256. 10. Robinson BW and Wilson DS. 1998. Optimal foraging, specialization, and a solution to Liem's paradox. Am Nat 151: 223–235. 11. Ungar PS. 2007. Dental Functional Morphology. In: Ungar P. (ed.), Evolution of the human diet: The Known, the Unknown, and the Unknowable. Oxford University Press, New York, pp. 39-55. 12. Deane AS. 2009. Early Miocene catarrhine dietary behaviour: the influence of the Red Queen Effect on incisor shape and curvature. Journal of Human Evolution. 56(3):275-285. 13. Scott RS, Ungar PS, Bergstrom TS, Brown CA, Grine FE, Teaford MF, and Walker A. 2005. Dental microwear texture analysis shows within-species diet variability in fossil hominins. Nature. 436:693-695. 14. Ungar PS, Merceron G, and Scott, RS. 2007. Dental Microwear Texture Analysis of Varswater Bovids and Early Pliocene Paleoenvironments of Langebaanweg, Western Cape Province, South Africa. J Mammal Evol. 14:163-181. 15. Prideaux GJ, Ayliffe LK, DeSantis LRG, Schubert BW, Murray PF, Gagan MK, and Cerling TE. 2009. Extinction implications of a chenopod browse diet for a giant Pleistocene kangaroo PNAS. 106(28):11646-11650. 16. Schubert BW, Ungar PS, Sponheimer M, and Reed KE. 2006. Microwear evidence for Plio-Pleistocene bovid diets from Makapansgat Limeworks Cave, South Africa. Paleogeography Paleoclimatology, Paleoecology. 241(2):301-319. 17. Scott JR, Godfrey LR, Jungers WL, Scott RS, Simmons EL, Teaford MF, Ungar PS, Walker A. 2009. Dental microwear texture analysis of two families of subfossil lemurs from Madagascar. J Hum Evol. 56:405-416. 18. Ungar PS, Teaford MF, Kay RF. 2004. Molar Microwear and Shearing Crest Development in Miocene Catarrhines. Anthropologie. 42(1):21-35. 19. Teaford MF, Oyen OJ. 1989. In vivo and in vitro turnover in dental microwear. Am J Phys Anthrop 80: 447-460. 20. Scott RS, Teaford MF, and Ungar, PS. 2012. Dental microwear texture and anthropoid diets. Am J Phys Anthrop. 147(4):551-579. 21. Ungar PS, Brown CA, Bergstrom TS, and Walker A. 2003. Quantification of Dental Microwear by Tandem Scanning Confocal Microscopy and Scale-Sensitive Fractal Analyses. Scanning 25(4):185-193. 22. Scott RS, Ungar PS, Bergstrom TS Brown CA, Childs, BE, Teaford MF, Walker A. 2006. Dental microwear texture analysis: technical considerations. Journal of Human Evolution. 51(4):339-349. 23. Grine FE. 1986. Dental evidence for dietary differences in Australopithecus and Paranthropus: a quantitative analysis of permanent molar microwear. J. Hum. Evol. 15:783-822.. 24. Ungar PS, Scott JR, Curran, SC, Dunsworth HM, Harcourt-Smith WEH, Lehmann T, Manthi FK, McNulty KP. 2012. Early Neogene environments in East Africa: Evidence from dental microwear of tragulids. Paleogeography, Paleoclimatology, Paleoecology. 342-343:84-96.

Figure 7 -- Microwear texture variables of fossils (bold) and extant primates. Outliers indicated by 9. Extreme values indicated by *.



CONCLUSIONS

• Here we present results of the first dental microwear texture analysis performed exclusively within African Miocene non-cercopithecoid catarrhines. Results corroborate previous studies in revealing no significant differences found in the microwear patterns among any of the fossil taxa tested herein ¹⁸.

• Our results suggest that despite differences in body size and dental complexity, the non-cercopithecoid primates of the early-mid African Miocene were consuming foods that had similar mechanical properties, at least for available individuals shortly before death ²³, which therefore caused similar microwear patterns.

• Obscuration of microwear by exogenous grit is unlikely, as fossil tragulids from the same deposits exhibit clear species differences in their microwear textures ²⁴.

• The microwear signatures of these fossil primates does not indicate that they were consuming identical foods, merely that they were eating mechanically similar types of foods, and plants with similar phytolith or dietary grit content, in the weeks before death.